Myostatin as a potential target for treatment of muscle contractures

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Overview

Neonatal Brachial Plexus Injury (NBPI)

- Most common cause of upper limb paralysis in childhood, occurring in 1.5 of every 1,000 live births. - Leads to the secondary formation of muscle contractures, or "limb stiffness."
- Contractures severely impede range of motion of the involved limbs, thereby impairing activities of daily living, and ultimately resulting in skeletal deformity and dysfunction.
- Current treatments for contractures are ineffective in restoring muscle function and joint range of motion. - To develop effective strategies for preventing and treating contractures, we first need to establish greater insights on contracture pathogenesis.

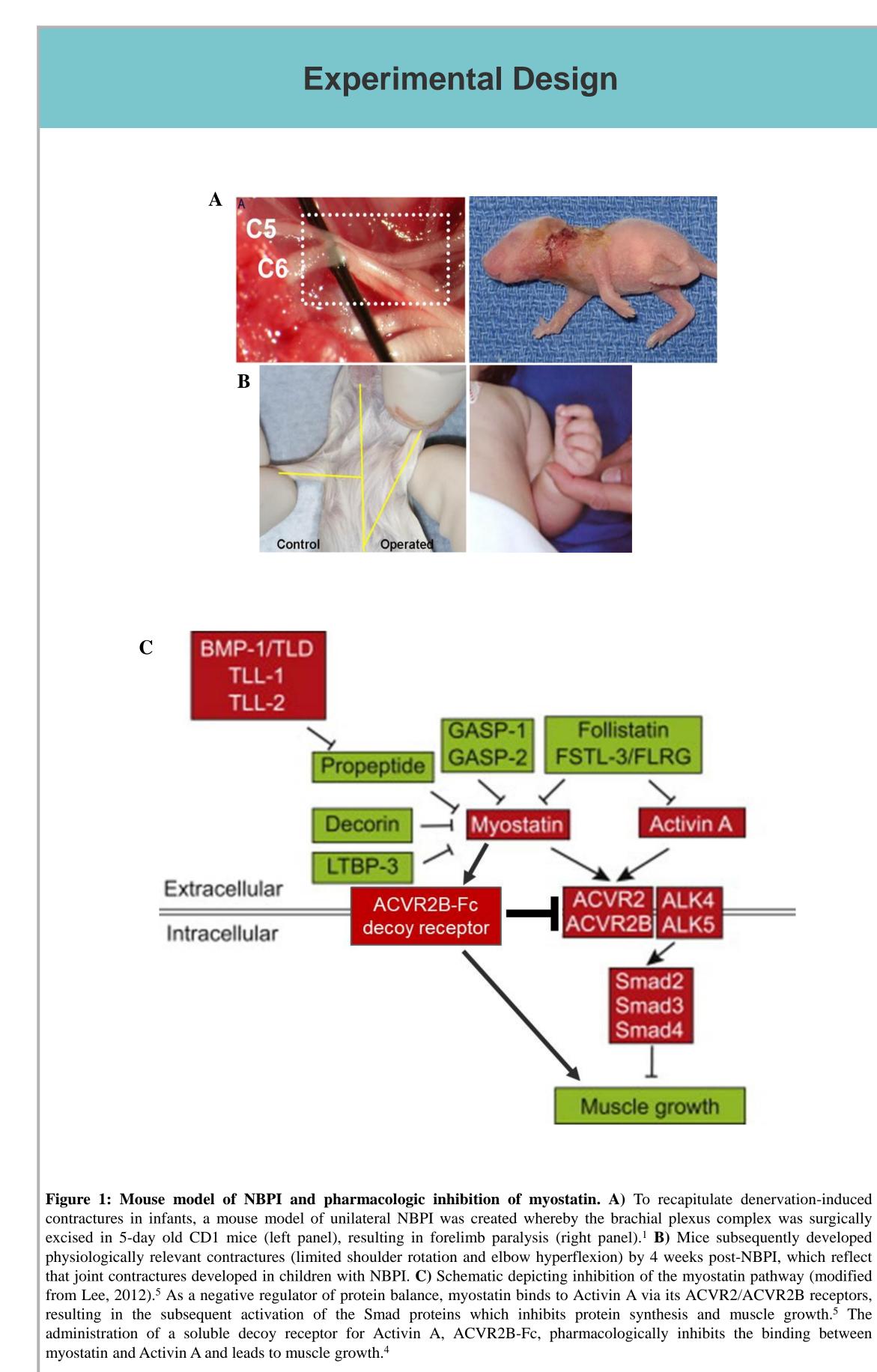
Contracture Pathogenesis

- Contractures following NBPI result from impaired longitudinal growth of the denervated muscle.¹ - Deficits in longitudinal growth of denervated muscles are driven by increased levels of proteasomemediated protein degradation, suggesting a dysregulation of protein balance in play.² - Proteasome inhibitors markedly reduce contracture formation but also block protein degradation
- nonspecifically, and prolonged treatment results in potential cumulative toxicity.³ * Hence, we need to identify safer strategies for preventing contractures by targeting muscle-specific regulators of the protein balance.

Myostatin Inhibition

- Myostatin is skeletal muscle-specific negative regulator of protein balance, and limits excess muscle growth by inhibiting protein synthesis.
- Pharmacologic inhibition of myostatin signaling dramatically enhances muscle growth in adult mice.⁴ - In this current study, we report that neonatal myostatin inhibition reduces NBPI-induced contracture

formation and preserves longitudinal growth of denervated muscles only in female mice. * <u>Our findings therefore reveal a sex-dependent role of myostatin signaling in the development of muscle</u> contractures after neonatal denervation injury.



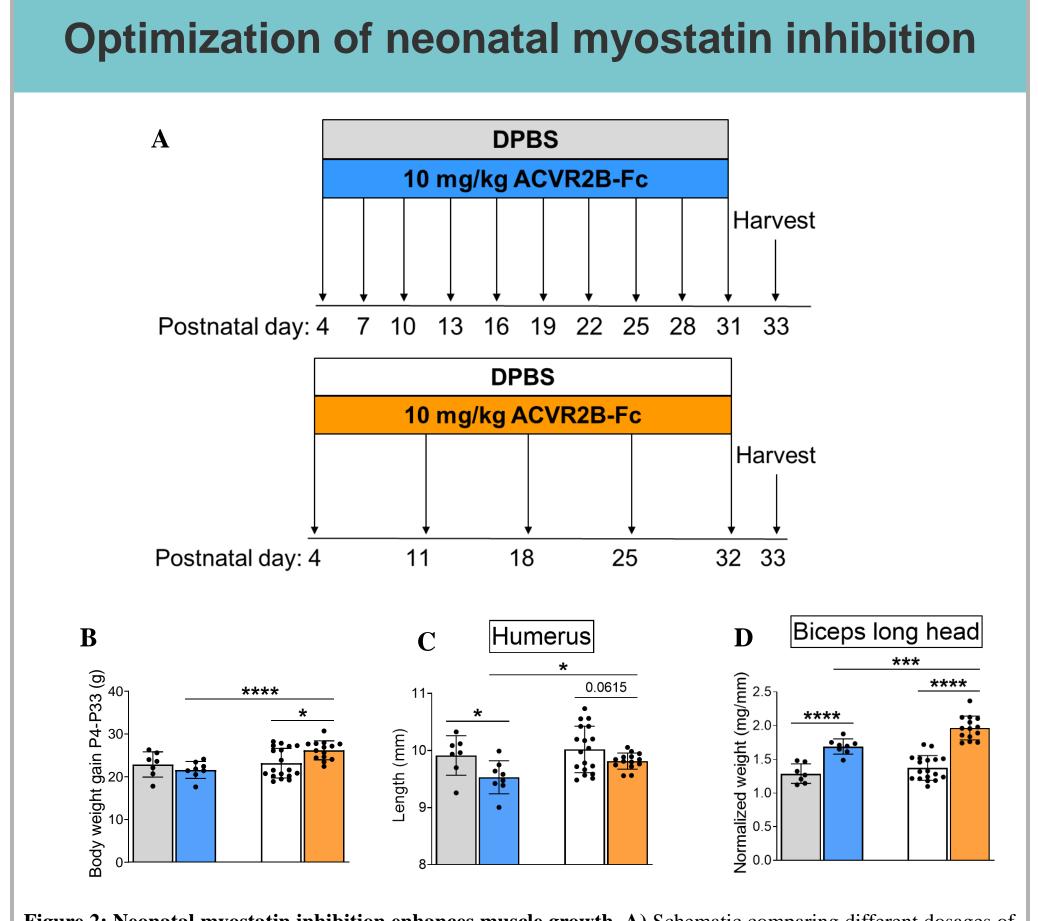


Figure 2: Neonatal myostatin inhibition enhances muscle growth. A) Schematic comparing different dosages of ACVR2B-Fc. The top schematic depicts administration of ACVR2B-Fc every three days, the dosage used in a previous study conducted on adult mice.⁴ The bottom schematic depicts weekly administration of ACVR2B-Fc. **B**) Treatment with ACVR2B-Fc every three days was less effective in promoting body weight gain than weekly treatment, and C) even reduced skeletal growth of the humerus. D) While both dosages increased the weight of biceps muscles, weekly treatment of ACVR2B-Fc resulted in greater skeletal muscle growth. All data are presented as mean \pm s.d. n = 7-18. Statistical analyses: (**B**), (**C**), (**D**) unpaired two-tailed Student's t-tests. The degree of significance between data sets is depicted as follows: *P<0.05, ***P<0.001, ****P<0.0001.

Sex-Specific Response to Neonatal Myostatin Inhibition – Developmental Growth

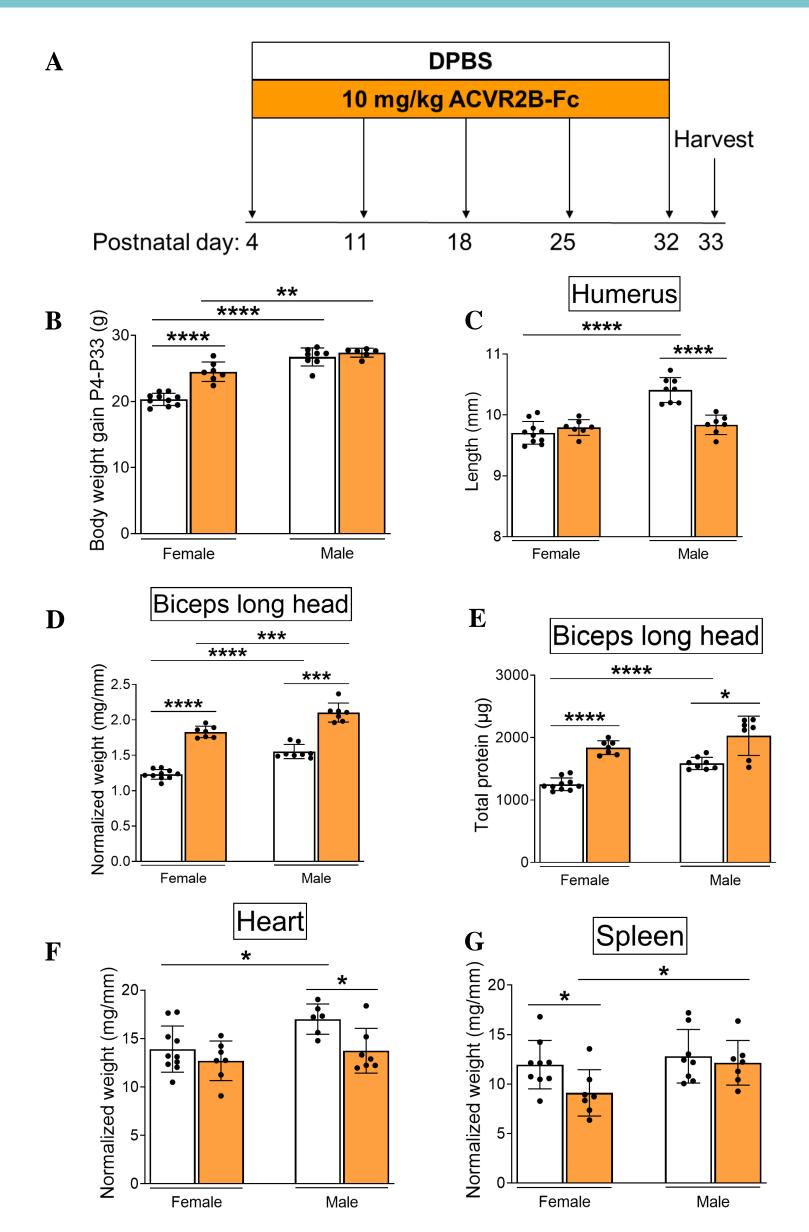
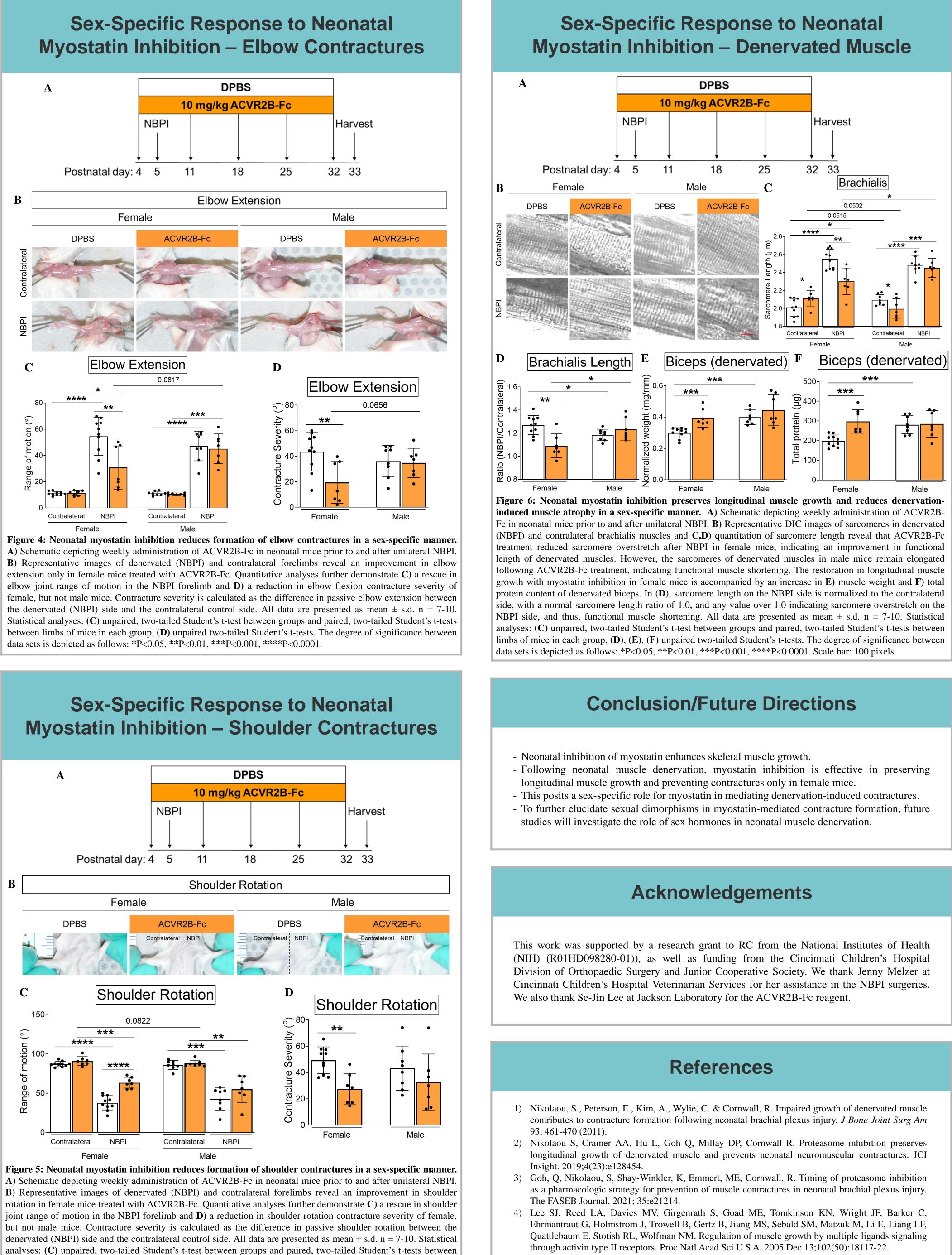


Figure 3: Sexual dimorphisms in developmental growth with neonatal myostatin inhibition. A) Schematic depicting weekly administration of ACVR2B-Fc in neonatal mice. B) Myostatin inhibition increased body weight in female mice but C) decreased humerus length in male mice. D) Myostatin inhibition increased the weight and E) total protein content of biceps muscles in both sexes. However, this increase in skeletal muscle growth was associated with sex-specific perturbations in non-skeletal muscle tissues, specifically reductions in F) hearts of male mice and G) spleens of female mice. All data are presented as mean \pm s.d. n = 6-10. Statistical analyses: (B), (C), (D), (E), (F), (G) unpaired two-tailed Student's t-tests. The degree of significance between data sets is depicted as follows: *P<0.05, **P<0.01, ***P<0.001 ****P<0.0001.





limbs of mice in each group, (**D**) unpaired two-tailed Student's t-tests. The degree of significance between data sets is depicted as follows: **P<0.01, ***P<0.001, ****P<0.0001.

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5) Lee SJ, Chapter 79 - Myostatin: Regulation, Function, and Therapeutic Applications, Editor(s): Joseph A. Hill, Eric N. Olson, Muscle, Academic Press, 2012, Pages 1077-1084.